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# Selective detection of pyrophosphate by new tripodal amine calix[4]arene-based Cu(II) complexes using indicator displacement strategy

Sarayut Watchasit <sup>a</sup>, Arpadsara Kaowliew <sup>a</sup>, Chomchai Suksai <sup>a,</sup>\*, Thawatchai Tuntulani <sup>b</sup>, Wittaya Ngeontae <sup>c</sup>, Chaveng Pakawatchai <sup>d</sup>

a Department of Chemistry and Center for Innovation in Chemistry, Faculty of Science, Burapha University, Chonburi 20131, Thailand

b Department of Chemistry, Faculty of Science, Chulalongkorn University, Bangkok 10330, Thailand

 $c$ Department of Chemistry and Center for Innovation in Chemistry, Faculty of Science, Khon Kean University, Khon Kean 40002, Thailand

<sup>d</sup> Department of Chemistry and Center for Innovation in Chemistry, Faculty of Science, Prince of Songkla University, Songkhla 90112, Thailand

# article info

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## **ABSTRACT**

Mono- and dinuclear Cu(II) complexes of p-tert-butylcalix[4]arene (CuL1 and CuL2, respectively) were synthesized, and their anion recognition abilities were explored. Recognition is efficiently signaled through the displacement of pyrocatechol violet bound to the receptor. For CuL2, recognition selectivity is ascribed to the tuning of the distance between donor atoms of anion guests and their ability to encompass the Cu<sup>2+</sup>–Cu<sup>2+</sup> distance within the cleft of **CuL2**. In addition, the preorganization of calix[4]arene in the cone conformation and steric hindrance of two bulky tripodal amine moieties are important factors in controlling the  $Cu^{2+}-Cu^{2+}$  distance. These factors caused **CuL2** to recognize pyrophosphate selectively with respect to other inorganic anions in 80/20 (v/v%) MeCN/H<sub>2</sub>O solution buffered with 10 mM HEPES at pH 6.4.

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Significant attention has been given to the development of anion sensing by indicator displacement assays  $(IDAs)^1$  $(IDAs)^1$ . This method is a simple, convenient, and increasingly popular approach to naked-eye anion sensors because an indicator is bound to a receptor by non-covalent interactions. IDAs rely upon competition between the indicator and the analyte in the host cavity. Consequently, a receptor is designed to bind a target analyte with a desired affinity, and an indicator must have a weaker affinity with the receptor than the analyte. Importantly, the indicator must absorb or emit light differently upon binding to the host and being in free form in solution.

Generally, anion recognition in the aqueous system is very challenging due to the strong hydration effects of anions. The utilization of a metal complex as a binding site for anions has been found to be the most successful strategy.<sup>[2](#page-3-0)</sup> Therefore, metal com-plexes are often used as IDA receptors.<sup>[3](#page-3-0)</sup> Normally, metal-bound ligands can bind anions more efficiently than water, allowing the detection of anions in aqueous solution. The metal center must have an unsaturated coordination sphere to accommodate the incoming anion guest.

IDA receptors for pyrophosphate  $(P_2O_7^{4-}$ , PPi), the product of ATP hydrolysis and involved in DNA polymerization in biological reactions,<sup>4</sup> have been developed by many research groups utilizing dinuclear zinc complexes of phosphotriesterase enzyme as the receptor module.<sup>[5](#page-3-0)</sup> Recently, Hong and Fabbrizzi reported that dinuclear  $Cu^{2+}$ –DPA complexes can be employed as PPi fluorescence sensors using the IDA concept. $<sup>6</sup>$  The metal–metal distance</sup> is found to play a key role in analyte preference. In order to obtain the selectivity toward PPi over other anions, especially phosphate anion ( $PO<sub>4</sub><sup>3–</sup>$ ), the metal–metal distance should not be less than 3.4 Å which is the Zn–Zn distance in phosphotriesterase.<sup>[7](#page-3-0)</sup>

Our group is currently working on the synthesis of calix[4]arenes containing tripodal amine as the recognition unit for use as ionophores for ion selective electrodes (ISEs). The rigidity of calix[4]arenes in the cone conformation plays an important role by providing a specific cavity to recognize specific guest molecules, due to preorganization of its skeleton.<sup>8</sup> The transannular distances in the lower rim of original calix[4]arene and its derivatives are in the range of 3.74-4.20  $\AA$ <sup>[9](#page-3-0)</sup> which are longer than the Zn-Zn distance in the dinuclear zinc enzyme. Therefore, calix[4]arenes may be a suitable building block for PPi using IDA strategies. In addition, the side arms attached to the calix[4]arene can control the size and shape of the recognition cavity of calix $[4]$ arene derivatives.<sup>[10](#page-3-0)</sup>

In this work, calix[4]arenes containing a tripodal amine have been chosen as IDA receptors for PPi. Furthermore, we expected that steric interactions between the tripodal amine and the rigidity of the calix[4]arene framework would play crucial roles in optimizing the metal–metal distance and provide more selectivity toward PPi. Herein, we report the synthesis and characterization of ionophores based on calix[4]arenes L1 and L2 and their mononuclear and dinuclear complexes with CuCl<sub>2</sub> (CuL1 and CuL2). We also





<sup>\*</sup> Corresponding author. Tel.: +66 038 103067; fax: +66 038 393494. E-mail address: [jomjai@buu.ac.th](mailto:jomjai@buu.ac.th) (C. Suksai).

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demonstrate that the dinuclear complex, CuL2, is a suitable receptor for indicator displacement assay of PPi.

Ionophores L1 and L2 were synthesized in two steps according to Scheme 1. The mono-calixaldehyde<sup>[11](#page-3-0)</sup> and bis-calixaldehyde<sup>[12](#page-3-0)</sup> (for L1 and L2, respectively) were reacted with 2-[bis(2-pyridyl $methyl$ )aminomethyl]aniline<sup>[13](#page-3-0)</sup> in dichloromethane, followed by in situ reduction with NaBH<sub>4</sub> in methanol to yield **L1** in 12% and L2 in 20%, respectively. The HRMS-ESI spectra of L1 and L2 show parent peaks at  $m/z$  1085.6671 and 1521.8715 assigned to the molecular ions [L+H]<sup>+</sup> and provide evidence that the calix[4]arene derivatives are the  $1:1^{14}$  $1:1^{14}$  $1:1^{14}$  and  $1:2^{15}$  condensation products. The two calix[4]arene derivatives are in the cone conformation, as supported by their NMR spectra (Figs. S1–S4, Supplementary data). The  $^1\mathrm{H}$  NMR spectrum of **L1** was consistent with an asymmetric calix[4]arene structure. In particular, two pairs of doublets for the protons of the methylene bridges were observed.

Two OH singlets were observed for L1 at low field, 9.47 and 10.18 ppm (ratio 2:1). These strong downfield shifts for the OH protons are indicative of a circular hydrogen bond at the lower rim of these derivatives, in agreement with the results reported by Frkanec et al. $^{16}$  The  $^1\mathrm{H}$  NMR spectrum of **L2** features a pair of doublets at 3.32 ppm and 4.43 ppm corresponding to the equatorial and the axial protons of the methylene bridging groups, respectively. We deduce that the cone structure is the major conformation of L1 and L2 in solution.

Addition of CuCl<sub>2</sub> to methanolic solutions of  $L1$  and  $L2$  gave green complexes of CuL1 and CuL2 in 72%<sup>[17](#page-3-0)</sup> and 42%<sup>18</sup> yields, respectively. The mass spectrum of **CuL1** shows the parent peak at m/z 1182.5424 which is assigned to the molecular ion of the mononuclear complex [CuL1Cl]<sup>+</sup>. For **CuL2**, the parent peak at  $m/z$  1751.6365 corresponds to the molecular ion of the dinuclear complex  $[Cu<sub>2</sub> L2Cl<sub>3</sub>]<sup>+</sup>$ . A crystal of **CuL1** was obtained upon slow evaporation of a methanolic solution and the structure was deter-mined by X-ray crystallography, [Figure 1](#page-2-0).<sup>[19](#page-4-0)</sup> It is clearly seen that the calixarene skeleton adopts a cone conformation. It should be noted that the phenolic hydrogen atoms are involved in strong intramolecular  $O-H \cdots O$  hydrogen bonding with the neighboring oxygen atoms to stabilize the cone conformation, in agreement with the solution structure deduced from the  ${}^{1}$ H NMR spectrum.

The crystal structure of CuL1 also shows that two Cu(II) centers coordinate with four nitrogen donors from the tripodal amine unit and two chloride bridging ligands to give a distorted octahedral geometry. The substantial elongation of the axial Cu1–Cl1\_2 and Cu1–N1 bonds [2.984 and 2.553(4) Å, respectively] compared to the equatorial Cu1–Cl1, Cu1–N2, Cu1–N3, and Cu1–N4  $[2.278(12), 2.066(3), 1.977(4),$  and  $1.994(4)$  Å, respectively is caused by the active Jahn–Teller distortion of the  $Cu<sup>2+</sup>$  ion. Interestingly, the mass spectrum of the CuL1 complex suggests that it is a mononuclear complex in solution. This implies that the mononuclear complex of CuL1 is the most stable species in solution while the dinuclear complex is the most stable species in the solid state.

In light of the crystal structure of **CuL1**, we expect that the dye and anions might occupy the bimetallic cleft of dinuclear complex CuL2. In this work, we chose pyrocatechol violet (PV) as a



Scheme 1. Synthetic procedures for L1, L2, CuL1, and CuL2. Reagents and conditions: (i) anhyd CH<sub>2</sub>Cl<sub>2</sub>, anhyd MgSO<sub>4</sub>, rt; (ii) NaBH<sub>4</sub>, CH<sub>3</sub>OH, reflux; (iii) CuCl<sub>2</sub>, MeOH, rt.

<span id="page-2-0"></span>

Figure 1. (a) ORTEP representation of the solid state structure of the dinuclear complex of CuL1 with two bridging chloride ligands and (b) the coordination environment of  $Cu<sup>2+</sup>$  in the complex. Thermal ellipsoids are drawn at 50% probability level (CCDC 767461).

competitive indicator.<sup>[20](#page-4-0)</sup> The yellow solution of PV was prepared in 80% acetonitrile aqueous solution buffered with 10 mM HEPES pH 6.4, which was then titrated with increasing amounts of CuL2 using the same solvent at 25  $\degree$ C. It was found that addition of CuL2 led to the disappearance of the absorption band of PV at 430 nm, with the simultaneous appearance of a new band at 670 nm and a color change from yellow to green (Fig. 2a). In addition, an isosbestic point was found at 488 nm, suggesting the presence of two equilibrium species. A Job plot (at 670 nm) was also obtained and suggested that the complex between CuL2 and PV was formed with a 1:1 stoichiometry (inset of Fig. 2a). Using the Benesi–Hildebrand method, the association constant  $(K_a)$  between PV and **CuL2** was found to be  $1.30 \times 10^4$  M<sup>-1</sup>.<sup>[21](#page-4-0)</sup>

Upon addition of various anions (as tetrabutylammonium salts, 3 equiv) to the ensemble [CuL2PV] solutions, only PPi was able to turn the color from green to yellow of the unbound dye, while other anions did not give rise to UV–vis spectral changes (Fig. 2b) or any color changes (Fig. 2c). Moreover, we also carried out displacement of PV from the **CuL2** cavity by phosphate containing biomolecules (AMP, ADP, and ATP). Results showed that both ADP and ATP were able to displace PV from the cleft of CuL2, whereas AMP was not (Fig. S5, Supplementary data). Therefore, CuL2 possessed high selectivity toward PPi over other anions. We tried to change the dye from PV to fluorescein. However, results of this ensemble did not show specific selectivity to any anions (Fig. S6, Supplementary data).

Titrating PPi with an ensemble solution [CuL2-PV] caused an absorbance increase around 430 nm and an absorbance decrease around 670 nm (hypsochromic shift), with a color change to yellow, revealing that the indicator was displaced from the cleft of CuL2 by the analyte [\(Fig. 3\)](#page-3-0). The UV–vis spectrum at 670 nm was completely saturated at 1.5 equiv of PPi. The binding constant between CuL2 and PPi was estimated by the competitive spectrophotometric method<sup>22</sup> and found to be 5.2  $\times$  10<sup>5</sup> M<sup>-1</sup>. The electrospray ionization mass spectrum (positive mode) of **CuL2** complex with PPi showed a molecular ion peak at  $m/z = 1824.57$  (Fig. S7, Supplementary data). The result thus confirmed the 1:1 complex species of CuL2:PPi.

Xu et al. have characterized crystallographically a ternary system complex of PPi with a mononuclear  $Cu^{2+}$  ion and a 2,2'-dipyridylamine (hdpa) ligand,  $[Cu(dhpa)]^{2+.23}$  $[Cu(dhpa)]^{2+.23}$  $[Cu(dhpa)]^{2+.23}$  In this case, one PPi anion acted as the bridging ligand to bring two units of [Cu(dhpa)] together, forming a dinuclear complex. Simultaneously, two PPi ions coordinating to the oxygen atoms of a discrete dinuclear complex also acted as the bridging atoms to hold those two discrete dimeric species together to form a tetranuclear complex. Compared to our system, we assume that PPi is bound within the bimetallic cleft of CuL2. Two oxygen atoms on each phosphorus of PPi coordinated



Figure 2. (a) UV–vis spectra obtained by addition of CuL2 (400  $\mu$ M) to PV (20  $\mu$ M) solution; (b) UV–vis spectra obtained by addition of various anions (3 equiv of tetrabutylammonium salts) to an ensemble solution [CuL2-PV] (20  $\mu$ M); and (c) color changes of the ensemble [CuL2-PV] 20  $\mu$ M after addition of various anions (3 equiv of tetrabutylammonium salts). From left to right: CuL2, PV, [CuL2·PV], H<sub>2</sub>PO<sub>4</sub>-, AcO<sup>-</sup>, PPi, BzO<sup>-</sup>, I-, Br<sup>-</sup>, Cl<sup>-</sup>, and F<sup>-</sup>. All experiments were carried out in 80/20 (v/v) MeCN/H<sub>2</sub>O solution buffered with 10 mM HEPES at pH 6.4.

<span id="page-3-0"></span>

Figure 3. UV-vis spectra obtained by addition of PPi (400  $\mu$ M) to an ensemble solution of  $\left[ \text{CuL2-PV} \right]$  (20 µM) in 80/20 (v/v) MeCN/H<sub>2</sub>O solution buffered with 10 mM HEPES at pH 6.4.

through one  $Cu^{2+}$  ion, which was similar to the binding mode of PPi with the dinuclear  $DPA-2Zn^{2+}$  derivatives reported by Yoon and co-workers<sup>24</sup> and Hong and co-workers.<sup>5b</sup>

Similar experiments have been run with the mononuclear CuL1 complex. The results showed that in the presence of any anions in an ensemble solution of [CuL1-PV] the yellow solution of the unbound dye was not observed (Fig. S8, Supplementary data). This result strongly supports the fact that the cooperative action of two  $Cu<sup>2+</sup>$  ions in solution is required for selective sensing of PPi.

In conclusion, we have successfully synthesized mono- and dinuclear Cu(II) complexes of calix[4]arene containing a tripodal amine, CuL1 and CuL2. CuL2 was demonstrated to be a remarkable IDA receptor for PPi. A rationale to account for the selectivity of CuL2 toward PPi requires matching of the distance between the donor atoms of PPi with the  $Cu^{2+}-Cu^{2+}$  distance in the **CuL2** cavity. In addition, the preorganization of calix[4]arene in the cone conformation and steric hindrance between the two bulky tripodal amine parts are the most important factors controlling the  $Cu^{2+}-Cu^{2+}$  distance. This resulted in selective recognition of CuL2 toward PPi over other anions. Further studies are underway in our laboratory to prepare anion selective electrodes from CuL2.

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# Supplementary data

Supplementary data (additional  $^1$ H and  $^{13}$ C NMR spectra of L1 and L2, and displacement results of CuL1 with various anions are available) associated with this article can be found, in the online version, at [doi:10.1016/j.tetlet.2010.04.095.](http://dx.doi.org/10.1016/j.tetlet.2010.04.095)

#### References and notes

- 1. (a) Wiskur, S. L.; Ait-Haddou, H.; Lavigne, J. J.; Anslyn, E. V. Acc. Chem. Res. 2001, 34, 963–972; (b) Nguyen, B. T.; Anslyn, E. V. Coord. Chem. Rev. 2006, 250, 3118– 3127.
- 2. (a) Kruppa, M.; König, B. Chem. Rev. 2006, 106, 3520–3560; (b) Suksai, C.; Tuntulani, T. Chem. Soc. Rev. 2003, 32, 192–202; (c) Martinez-Manez, R.; Sancenon, F. Chem. Rev. 2003, 103, 4419–4476; (d) Suksai, C.; Tuntulani, T. Top. Curr. Chem. 2005, 255, 163–198; (e) Kruppa, M.; Mandl, C.; Miltschitzky, S.; König, B. J. Am. Chem. Soc. 2005, 127, 3362–3365; (f) Lim, M. H.; Wong, B. A.; Pitcock, W. H.; Mokshagundam, D.; Baik, M.-H.; Lippard, S. J. Am. Chem. Soc.

2006, 128, 14364–14373; (g) Salo, T. M.; Helaja, J.; Koskinen, A. M. P. Tetrahedron Lett. 2006, 47, 2977–2980; (h) Lee, D. H.; Im, J. H.; Son, S. U.; Chung, Y. K.; Hong, J.-I. J. Am. Chem. Soc. 2003, 125, 7752–7753; (i) Guo, Z.; Zhu, W.; Tian, H. Macromolecules 2010, 43, 739–744.

- 3. (a) Amendola, V.; Bergamaschi, G.; Buttafava, A.; Fabbrizi, L.; Monzani, E. J. Am. Chem. Soc. 2010, 132, 264–268; (b) Zhang, T.; Anslyn, E. A. Tetrahedron 2004, 60, 11117–11124; (c) Hanshaw, R. G.; Hilkert, S. M.; Jiang, H.; Smith, B. D. Tetrahedron Lett. 2004, 45, 8721–8724; (d) Zhang, T.; Anslyn, E. V. Org. Lett. 2007, 9, 1627–1629; (e) Swamy, K. M. K.; Kwon, S. K.; Lee, H. N.; Kumar, S. M. S.; Kim, J. S.; Yoon, J. Tetrahedron Lett. 2007, 48, 8683–8686; (f) Jang, H. H.; Yi, S.; Kim, M. H.; Kim, S.; Lee, N. H.; Han, M. S. Tetrahedron Lett. 2009, 50, 6241–6243; (g) Zhang, S.; Glass, T. E. Tetrahedron Lett. 2010, 51, 112–114; (h) Yin, C.; Huo, F.; Yang, P. Sens. Actuators, B 2005, 109, 291–299; (i) Fabbrizzi, L.; Marcotte, N.; Stomeo, F.; Taglietti, A. Angew. Chem., Int. Ed. 2002, 41, 3811–3814; (j) McDonough, M. J.; Reynolds, A. J.; Lee, W. Y. G.; Jolliffe, K. A. Chem. Commun. 2006, 2971–2973.
- 4. Ronaghi, M.; Karamohamed, S.; Pettersson, B.; Uhlén, M.; Nyrén, P. Anal. Biochem. 1996, 242, 84–89.
- 5. (a) Kim, S. K.; Lee, D. H.; Hong, J.-I.; Yoon, J. Acc. Chem. Res. 2009, 42, 23–31; (b) Lee, J. H.; Park, J.; Lah, M. S.; Chin, J.; Hong, J.-I. Org. Lett. 2007, 9, 3729–3731; (c) Morgan, P. B.; He, S.; Smith, R. C. Inorg. Chem. 2007, 46, 9262–9266; (d) Cho, H. K.; Lee, D. H.; Hong, J.-I. Chem. Commun. 2005, 1690–1692; (e) Mangalum, A.; Smith, R. C. Tetrahedron 2009, 65, 4298–4303; (f) Lee, D. H.; Kim, S. Y.; Hong, J.- I. Tetrahedron Lett. 2007, 48, 4477–4480.
- 6. (a) Hong, J.-I.; Kim, S. Y. Tetrahedron Lett. 2009, 50, 1951–1953; (b) Fabbrizzi, L.; Marcotte, N.; Stomeo, F.; Taglietti, A. Angew. Chem., Int. Ed. 2002, 41, 3811-3814.
- 7. Benning, M. M.; Shim, H.; Raushel, F. M.; Holden, H. M. Biochemistry 2001, 40, 2712–2722.
- 8. (a) Gutsche, C. D.; Reddy, P. A. J. Org. Chem. 1991, 56, 4783–4791; (b) Dijkstra, P. J.; Brunink, J. A. J.; Bugge, K.-E.; Reinhoudt, D. N.; Harkema, S.; Ungaro, R.; Ugozzoli, F.; Ghidini, E. J. Am. Chem. Soc. 1989, 111, 7567–7575; (c) Froidevaux, P.; Harrowfield, J. M.; Sobolev, A. N. Inorg. Chem. 2000, 39, 4678–4687.
- 9. Lipkowitz, K. B.; Pearl, G. J. Org. Chem. 1993, 58, 6729–6736.
- 10. (a) Scheeder, J.; van Duynhoven, J. P. M.; Engbersen, J. F. J.; Reinhoudt, D. N. Angew. Chem., Int. Ed. 1996, 35, 1092–1093; (b) Kim, S. K.; Kim, S. H.; Kim, H. J.; Lee, S. H.; Ko, J.; Bartsch, R. A.; Kim, J. S. Inorg. Chem. 2005, 44, 7866–7875; (c) Chang, K.-C.; Su, I. –H.; Senthilvelan, A.; Chung, W.-S. Org. Lett. 2007, 9, 3363– 3366.
- 11. Groenen, L. C.; Ruël, B. H. M.; Casnati, A.; Verboom, W.; Pochini, A.; Ungaro, R.; Reinhoudt, D. N. Tetrahedron 1991, 47, 8379–8384.
- 12. Navakun, K.; Tuntulani, T.; Ruangpornvisuti, V. J. Inclusion Phenom. 2000, 38, 113–122.
- 13. Burdette, S. C.; Frederickson, C. J.; Bu, W.; Lippard, S. J. J. Am. Chem. Soc. 2003, 125, 1778–1787.
- 14. Ionophore L1, <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, ppm):  $\delta$  10.20 (s, 1H, -OH), 9.49 (s, 2H  $-OH$ ), 8.44 (d, 2H, J = 4.0 Hz, ArH), 7.49 (t, 2H, J = 6.0 Hz, ArH), 7.37 (d, 2H,  $J = 4.0$  Hz, ArH), 7.34 (d, 2H,  $J = 8.0$  Hz, ArH), 7.13 (s, 2H, ArH), 7.12 (s, 1H, ArH), 7.07 (d, 2H, J = 2.0 Hz, ArH), 7.05 (s, 2H, ArH), 7.03 (s, 2H, ArH), 7.02 (s, 2H, ArH), 6.94 (m, 2H, ArH), 8.82 (s, 1H,  $-NH-$ ), 6.54 (t, 1H,  $J = 7.6$  Hz, ArH), 6.40 (d, 1H, J = 8.0 Hz, ArH), 4.70 (d, 2H, J = 3.2 Hz, -CH<sub>2</sub>-NH-), 4.64 (s, 4H, -O-CH<sub>2</sub>-O-)<br>4.53 (d, 2H, J = 12.8 Hz, Ar-CH<sub>2</sub>-Ar), 4.24 (d, 2H, J = 13.6 Hz, Ar-CH<sub>2</sub>-Ar), 3.83  $(s, 4H, -CH_2-N)$ , 3.69  $(s, 2H, -CH_2-N)$ , 3.41  $(dd, 4H, J = 7.6 Hz, J = 12.8, 13.6 Hz$ , Ar–CH<sub>2</sub>–Ar), 1.23 (s, 36H, p-tert-butyl); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 159.24, 156.11, 149.12, 149.04, 148.39, 148.29, 148.09, 147.85, 143.50, 143.12, 136.30, 133.61, 130.94, 128.74, 128.53, 128.12, 128.07, 127.75, 127.69, 126.56, 125.82, 125.71, 125.66, 123.18, 121.90, 121.51, 121.10, 115.32, 110.80, 110.20, 74.78, 60.13, 58.47, 41.73, 34.26, 33.99, 33.93, 33.00, 32.18, 31.50, 31.25; HRMS-ESI: [M+H]<sup>+</sup> calcd for C<sub>72</sub>H<sub>84</sub>N<sub>4</sub>O<sub>5</sub>, 1085.6670: found 1085.6671.
- 15. Ionophore L2, <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.42 (d, 4H, J = 4.0 Hz, ArH), 7.65 (d. 2H, J = 2.4 Hz, ArH), 7.46 (m, 4H, ArH), 7.35 (d, J = 7.6 Hz, 2H, ArH), 7.30 (s, 4H, ArH), 7.19 (m, 2H, ArH), 7.06 (m, 8H, ArH), 7.05 (m, 2H, ArH), 6.95 (d, 4H, J = 8.4 Hz, ArH), 6.89 (s, 4H, ArH) 6.86 (d, 2H, J = 7.2 Hz, ArH), 6.72 (s, 2H, –NH–), 6.54 (m, 2H, ArH), 6.44 (d, 2H, J = 8.0 Hz, ArH), 4.53 (d, 4H, J = 4.8 Hz, -CH<sub>2</sub>-NH–), 4.44 (d, 4H, J = 12.8 Hz, Ar–CH<sub>2</sub>–Ar), 4.35 (s, 8H, –CH<sub>2</sub>–O–), 3.80 (s, 8H, – CH<sub>2</sub>–N), 3.68 (s, 4H, –CH<sub>2</sub>–N), 3.33 (d, 4H, J = 13.2 Hz, Ar–CH<sub>2</sub>–Ar), 1.26 (s, 18H, *p*-tert-butyl), 1.03 (s, 18H, *p*-tert-butyl); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, ppm):  $\delta$ 159.23, 156.24, 150.35, 149.73, 149.07, 148.21, 147.34, 141.69, 136.29, 133.12, 130.97, 128.77, 128.56, 128.52, 127.93, 127.63, 125.72, 125.20, 123.20, 121.91, 121.55, 120.71, 115.35, 110.78, 110.32, 74.16, 66.65, 60.16, 58.47, 41.76, 34.04, 33.84, 31.83, 31.66, 31.10; HRMS-ESI:  $[M+H]^+$  calcd for C<sub>100</sub>H<sub>112</sub>N<sub>8</sub>O<sub>6</sub>, 1521.8705: found 1521.8715.
- 16. Frkanec, L.; Višnjevac, A.; Kojić-Prodić, B.; Žinić, M. Chem. Eur. J. 2000, 6, 442– 453.
- 17. CuL1; A methanolic solution of CuCl<sub>2</sub>.2H<sub>2</sub>O (24 mg, 0.14 mmol) in 5 mL CH<sub>3</sub>OH was added to a suspension of L1 (0.108 g, 0.09 mmol) in methanol giving a deep-green solution. The solution was allowed to stand at room temperature. After 1 week, green block-shaped X-ray diffraction quality single crystals of  $CuL1$  were obtained (80 mg, 73%). HRMS-ESI:  $[M+C1]^+$  calcd for **CuL1** were obtained (80 mg, 73%). HRMS-ESI:  $[M+C1]^+$  $C_{72}H_{84}ClCuN<sub>4</sub>O<sub>5</sub>$ , 1182.5426: found 1182.5424.
- 18. CuL2: A methanolic solution of CuCl<sub>2</sub>·2H<sub>2</sub>O (26 mg, 0.15 mmol) was added to a methanolic suspension of ionophore L2 (110.9 mg, 0.06 mmol); the color of the solution changed to deep-green immediately. After standing the green solution at room temperature for 1 week, the deep-green solid appeared. This was filtered and washed with MeOH to give CuL2 in 42% yield (111 mg). HRMS-ESI: [M+Cl]<sup>+</sup> calcd for C<sub>100</sub>H<sub>112</sub>Cl<sub>3</sub>Cu<sub>2</sub>N<sub>8</sub>O<sub>6</sub>, 1751.6362: found 1751.6365.
- <span id="page-4-0"></span>19. Crystal data for **CuL1**:  $C_{146}H_{180}Cl_2Cu_2N_8O_{14}$ , Mr = 2539.99, T = 293(2) K, triclinic, space group  $P\overline{1}$  a = 13.3699(10),  $\overline{b}$  = 15.5074(12), c = 17.9970(13) Å,  $\alpha$  = 94.853° (2),  $\beta$  = 90.013° (2),  $\gamma$  = 99.694° (2),  $V$  = 3664.5(5) Å<sup>3</sup>,  $\rho_{\text{calcd}}$  = 1.151 g cm<sup>-3</sup>,  $\mu$  = 0.422 mm<sup></sup>  $R1 = 0.0974$ , wR2 = 0.2936, R indices (all data):  $R1 = 0.1511$ , wR2 = 0.3344. Crystallographic data for CuL1 are available upon request from the Cambridge Crystallographic Data Base (CCDC 767461).
- 20. All spectrophotometric titrations were performed in 80/20 (v/v) MeCN/H<sub>2</sub>O solution buffered with 10 mM HEPES at pH 6.4, thermostated at 25 °C. Receptor/indicator affinity constants were determined by adding aliquots of a 400  $\mu$ M CuL2 complex solution to a 20  $\mu$ M solution of PV. After each addition,

the UV–vis absorption spectra of the indicator solution were recorded in a quartz cuvette. Similar titration experiments were performed with PPi. In typical titrations, aliquots of PPi (400  $\mu$ M) were added to an ensemble solution  $(20 \mu M)$  of  $[Cul2PV]$ .

- 21. Roy, P.; Dhara, K.; Manassero, M.; Ratha, J.; Banerjee, P. Inorg. Chem. 2007, 46, 6405–6412.
- 22. Niikura, K.; Bisson, A. P.; Anslyn, E. V. J. Chem. Soc., Perkin Trans. 2 1999, 1111-1114.
- 23. Xu, J.-Y.; Tian, J.-L.; Zhang, Q.-W.; Zhao, J.; Yan, S.-P.; Liao, D.-Z. Inorg. Chem. Commun. 2008, 11, 69–72.
- 24. Jang, Y. J.; Jun, E. J.; Lee, Y. J.; Kim, Y. S.; Kim, J. S.; Yoon, J. J. Org. Chem. 2005, 70, 9603–9606.